

15-ACETYLTHIOXY-FURODYSININ LACTONE, A POTENT LTB₄ RECEPTOR
PARTIAL AGONIST FROM A MARINE SPONGE OF THE GENUS DYSIDEA

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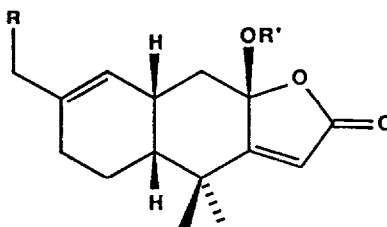
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Summary: A novel sesquiterpene thioacetate with high binding-affinity for the human leukotriene B₄ (LTB₄) receptor has been isolated from the sponge Dysidea sp. collected in Palau. The structure of the new compound was determined on the basis of spectral data and the compound was prepared semi-synthetically by photo-oxidation of the known metabolite, thiofurodysin acetate.

A decade ago, investigations of a marine sponge of the genus Dysidea from Australia yielded the furanoid sesquiterpenes furodysin and furodysin in along with their related thioacetates, which were the first reported naturally occurring terpene thiol derivatives¹. Since this original study, reports on the isolation of oxidation products of furodysin and furodysin in from the dorid nudibranchs Chromodoris funerea² and Hypselodoris zebra³ have appeared, but oxidation products of the related thioacetates were not reported. During our search for antagonists of leukotriene receptors, the crude extract of a Dysidea sp. sponge collected in Palau was shown to be active (IC₅₀ = 30 µg/mL) in a LTB₄ receptor binding assay⁴. This activity was shown to be due to furodysin in lactone thioacetate (1), the first reported oxidation product of thiofurodysin in acetate.

Dysidea sp. (specimen no. 81-057, 500 grams dry wt.) was collected by hand using SCUBA along the outer reef (-30 m) due west of Malakal harbor in the Palau Islands. The crude methanol extract was partitioned between water and CH₂Cl₂. The organic layer was chromatographed on silica using the LTB₄ radioligand binding assay⁴ to monitor biological activity, yielding 5.0 mg (0.01% dry wt) of 1 (mp 144-145°C; [α]^{CHCl₃} = -178°). The HRMS of 1 established the formula as C₁₇H₂₂O₄S [found m/z = 323.1305, C₁₇H₂₃O₄S(M+H) requires m/z = 323.1317] and the IR (KBr, 3300, 1744, 1693 cm⁻¹) suggested the presence of a thioacetate and a γ-hydroxy-γ-lactone. ¹H and ¹³C NMR data⁵ were almost identical with the published spectra of furodysin in lactone³ (2) and O-methylfurodysin in lactone² (3) with replacement of the vinylic methyl group signals with those indicative of a CH₂-S-Ac group. The foregoing data allowed assignment of the structure as 1, which was supported in turn by extensive 2D NMR analysis and verified by photo-oxidation of furodysin in thioacetate to furodysin in lactone thioacetate (1) in the presence of a polymer-bound rose bengal catalyst using previously published conditions². This semi-synthetic compound was identical in all respects to the natural product.

- 1: R = AcS, R' = H
 2: R = R' = H
 3: R = H, R' = Me



In an isolated U-937 cell membrane [^3H]LTB₄ receptor binding assay⁴, LTB₄ and furodysinin lactone thioacetate (1) have binding affinities (K_i) of 0.1 nM and 0.2 μM respectively. In a human lung membrane [^3H]LTB₄ receptor binding assay⁶, LTB₄ and 1 have binding affinity at 0.2 nM and 0.1 μM respectively. In differentiated human U-937 cells, furodysinin lactone thioacetate (1) exhibited partial agonist-like activity and induced LTB₄-receptor mediated Ca^{2+} mobilization with an ED_{50} of 4.0 μM ⁷. The maximal extent of the Ca^{2+} mobilization activity, however, was only 50-60% of that induced by 1 μM LTB₄. These results indicate that 1 could bind to human lung and U-937 cell membrane LTB₄ receptors and promote LTB₄ receptor mediated calcium mobilization in U-937 cells⁸.

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REFERENCES

1. Kazlauskas, R., Murphy, P.T., Wells, R.J., Daly, J.J. and Schonholzer, P., Tetrahedron Letters, 4951 (1978).
2. Carte', B., Kernan, M.R., Barrabee, E.B., Faulkner, D.J., Matsumoto, G.K. and Clardy, J., J. Org. Chem. 51, 3528 (1986).
3. Grode, S.H. and Cardellina, J.H., J. Nat. Prod., 47, 76 (1984).
4. Bomalski, J.S. and Mong, S., Prostaglandins 33, 855 (1987).
5. $^1\text{H-NMR}$ (CDCl_3) δ 1.24 (s, 3H), 1.41 (s, 3H), 1.55 (t, 1H, J = 13.5 HZ), 1.70 (2H, m), 2.04 (bd, 2H, J = 8 HZ), 2.32 (dd, 1H, J = 13.5, 3.6 HZ), 2.35 (s, 3H), 2.90 (m, 1H), 3.48 (s, 2H), 5.67 (bd, 1H, J = 8 HZ), 5.69 (s, 1H); $^{13}\text{C-NMR}$ (CDCl_3) δ 18.4 (t), 25.4 (q), 26.8 (q), 28.1 (t), 30.4 (d), 30.5 (q), 35.4 (t), 38.4 (s), 40.5 (t), 47.2 (d), 104.9 (s), 115.2 (d), 127.4 (d), 137.8 (s), 170.1 (s), 174.6 (s), 195.5 (s).
6. Lewis, M.A., Mong, S., Vessella, R.L. and Crooke, S.T., Biochem. Pharmacol., 34, 4311 (1985).
7. Mong, S., Miller, J., Wu, H-L. and Crooke, S.T., J. Pharmacology Exp. Therapeutics, 244, 508 (1988).
8. A QSAR study involving 10-15 furodysinin related compounds is in progress and will be published at a later date.

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